



# Role of adhesion receptor trafficking in 3D cell migration

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This review discusses recent advances in our understanding of adhesion receptor trafficking *in vitro*, and extrapolates them as far as what is currently possible towards an understanding of migration in three dimensions *in vivo*. Our specific focus is the mechanisms for endocytosis and recycling of the two major classes of cell-matrix adhesion receptors, integrins and syndecans. We review the signalling networks that are employed to regulate trafficking and conversely the effects of trafficking on signalling itself. We then define the contribution that this element of the migration process makes to processes such as wound healing and tumour invasion.

## Addresses

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## Introduction

The ability of cells to translocate *in vivo* is a fundamental requirement for embryonic development and tissue homeostasis: it also makes an essential contribution to the aetiology of, and host response to, virtually every disease condition. An understanding of the complex molecular mechanisms that enable cell migration would therefore generate insights into a diverse range of biological processes, as well as offer the prospect of modulating aberrant movement. It is understandable therefore that there has been intense interest in defining the modes of migration employed by cells *in vivo*. Using a diverse range of model systems, from cultured cells on two-dimensional surfaces to intravital examination of xenografts, apparently distinct phenotypic processes have been described, including lamellipodial migration in 2D,

and mesenchymal, amoeboid and lobopodial migration in 3D. In other articles in this issue, these different modes of migration are described in detail.

Regardless of mode, each type of cellular translocation shares some but not all, of the following features: receptor recognition of extracellular matrix (ECM) topography, formation and turnover of clustered adhesion signalling complexes, adaptation and deployment of dynamic cytoskeletal polymers, membrane uptake and delivery, and polarisation and spatial control of signalling. Whilst each of these features can be examined in isolation, it is likely that they are closely coordinated. Thus, adhesion complex clustering may be determined by ECM topography and/or cytoskeletal architecture, and membrane dynamics may control the sites where signalling occurs. A current aim is therefore to combine information obtained from highly reductionist approaches into high order models of migration.

## Integrins

Integrins are a major family of adhesion receptors. In mammals, 18  $\alpha$  and 8  $\beta$  integrin genes encode polypeptides that combine to form 24  $\alpha\beta$  heterodimeric receptors [1,2]. Both subunits are non-covalently associated, type I transmembrane proteins with large extracellular and mostly short cytoplasmic domains. The combined extracellular domains engage a range of extracellular matrix and cell surface ligands, whilst the cytoplasmic domains engage the actin cytoskeleton via a series of linker proteins [1,2]. Integrins enable cells to sample the topology and mechanochemical properties of their pericellular environment and respond by changing their position and differentiated state [1,2].

The regulation of integrin affinity by ligand and cytoskeletal proteins has been extensively studied, but in recent years the endocytic trafficking of integrins has emerged as a complementary mechanism through which the availability of integrins at the plasma membrane is controlled. Integrins are internalised via many of the best-characterised endocytic routes, and this dictates the ability of the receptors to promote cell migration in two and three dimensions (reviewed in [3,4]). For example, endocytosis controls the turnover of focal adhesions and therefore cell migration in 2D [5–7], whilst direct interactions between  $\alpha v\beta 6$  integrin and HAX-1 control receptor endocytosis, and have been shown to regulate invasion in 3D ECM [8].

Following endocytosis, integrins, like other cargo receptors, are sorted in early endosomes for degradation or recycling back to the plasma membrane [3,9,10]. As the

degradative turnover of integrins is relatively slow, endocytic recycling is considered to play a major role in regulating the spatiotemporal availability of integrins at the plasma membrane. In this context, several studies have demonstrated that the recycling of integrins contributes to adhesion complex formation and migration in 2D [3,11,12].

Accumulating evidence suggests that trafficking integrins also play an important role in regulating invasive migration in 3D [13,14]. Indeed, in cells migrating in 3D-microenvironments, the vesicular regulators that control integrin trafficking accumulate towards the invasive front [15,16,17\*] (Figure 1). It is notable that specific integrin heterodimers make different contributions to this process. For example,  $\alpha\text{v}\beta 3$  and  $\alpha 5\beta 1$  integrins bind to similar ligands, but can act antagonistically: whilst both integrins promote migration, they do so by eliciting different signalling responses and in fact mutually suppress each other [18]. Phosphorylation of rabaptin-5 by PKD promotes Rab4-dependent  $\alpha\text{v}\beta 3$  recycling, and this in turn promotes directionally persistent lamellipodial migration in 2D and invasion into 3D ECM in the absence of fibronectin (FN) [19,20\*\*]. However, in the presence of FN, this  $\alpha\text{v}\beta 3$ -recycling pathway suppresses invasive migration. This is because  $\alpha\text{v}\beta 3$ , and  $\alpha\text{v}\beta 3$  recycling, inhibit the recycling and pro-invasive activity of  $\alpha 5\beta 1$  [16,19,20\*\*]. When  $\alpha\text{v}\beta 3$  (or its recycling) is inhibited, or if cells express cancer-associated forms of mutant p53,  $\alpha 5\beta 1$  associates with the Rab11-effector Rab-coupling protein (RCP), and rapidly recycles to the plasma membrane to promote invasion into FN-rich ECM [16,20\*\*,21]. Production of phosphatidic acid by DGK $\alpha$  promotes the recruitment of RCP to the front of invasive cancer cells via its C2 domain, resulting in localised trafficking in this subcellular region [17\*]. Interestingly, RCP-driven  $\alpha 5\beta 1$  recycling does not influence

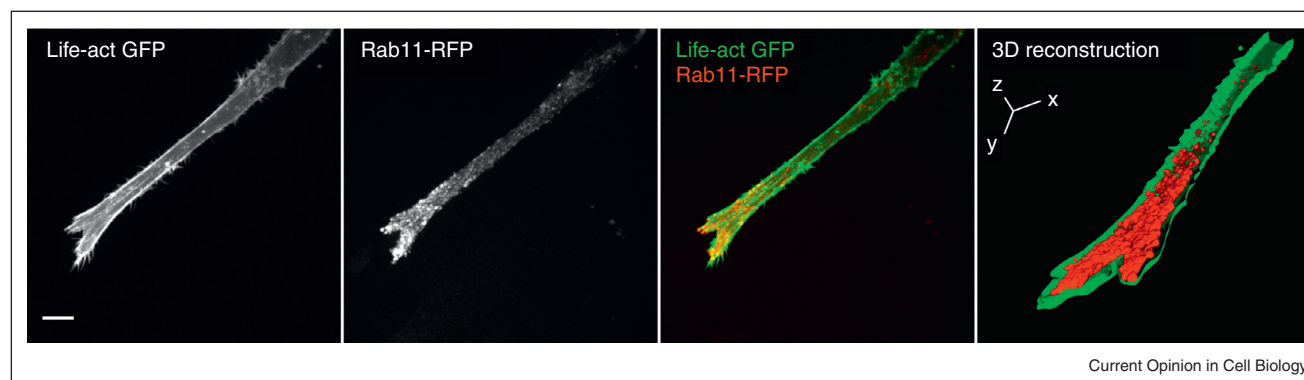
the ability of the integrin to mediate attachment via its ligand FN: instead,  $\alpha 5\beta 1$  and RCP recruit receptor tyrosine kinases and regulate their trafficking and signalling to promote invasion [16,21,22].

$\alpha 5\beta 1$  trafficking can also promote invasive migration in FN-rich ECM through Rab25, a Rab11 family member that is upregulated in aggressive ovarian cancer [23]. Rab25 binds directly to the cytoplasmic tail of  $\beta 1$  to direct  $\alpha 5\beta 1$  trafficking at the front of invading cells [15]. Here, endocytosed integrins are delivered to the Rab25 compartment at the cell front, and inactive integrins are trafficked directly back to the vicinal membrane [15]. Active  $\alpha 5\beta 1$  heterodimers are, however, trafficked via Rab25-positive late endosomes to lysosomes towards the rear of cell. Here, CLIC3, which is co-upregulated with Rab25 in a subset of aggressive ovarian and pancreatic cancers, promotes the recovery of  $\alpha 5\beta 1$  from lysosomes and recycling to the plasma membrane at the rear of the cell to facilitate invasion [24\*\*]. Thus, Rab25 can coordinate process extension, by recycling unligated integrin to the cell front, with retraction by recycling active integrins to the cell rear where they can promote signals for forward movement.

## Syndecans

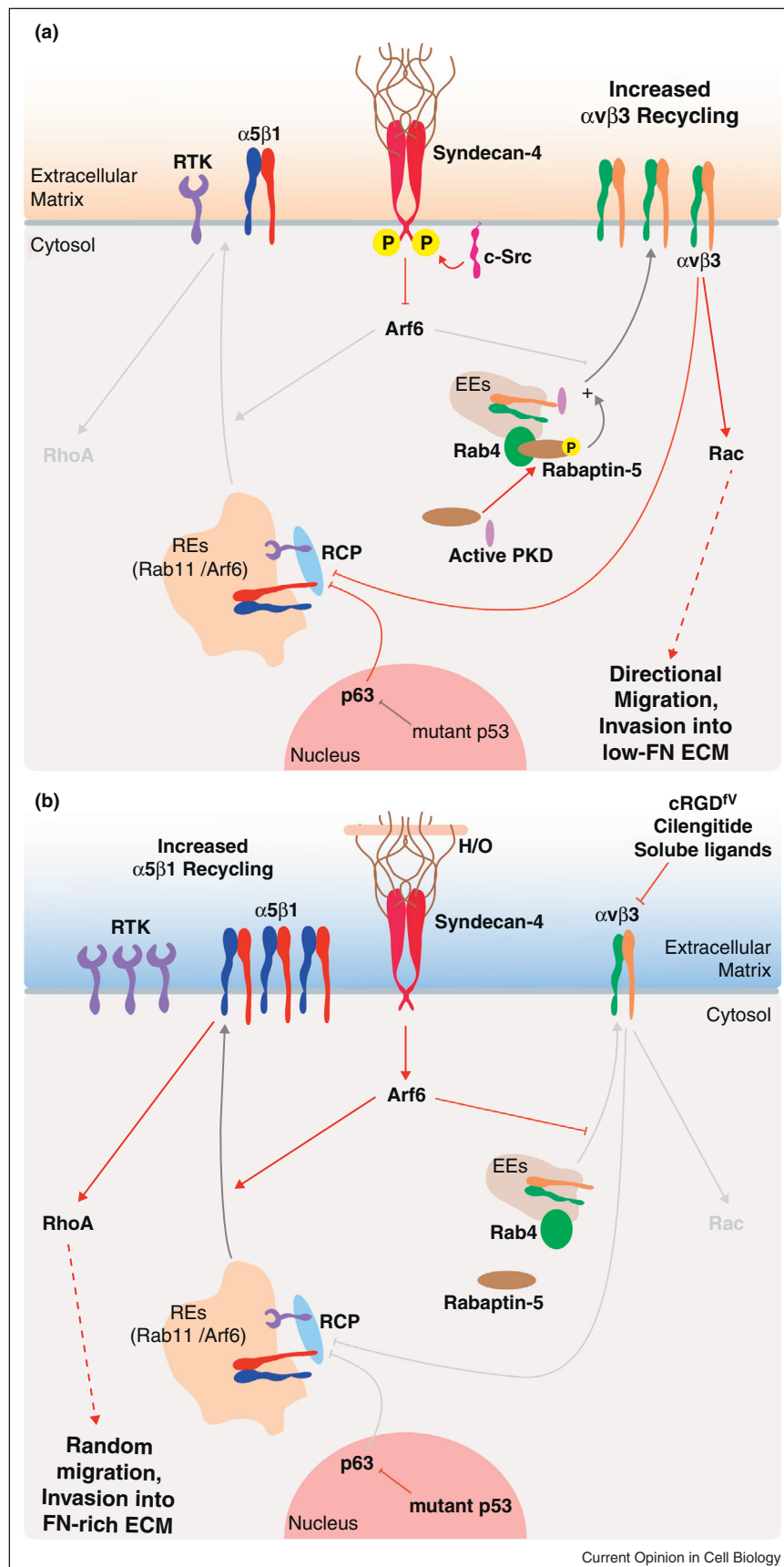
Syndecans are a small family of membrane-intercalated proteoglycans that serve as receptors for extracellular matrix ligands and growth factors [25]. In mammals, there are four members. It is remarkable that most ECM molecules possess both integrin-binding and syndecan-binding sites, and a clear synergistic relationship exists between these two families. For example, adhesion complex formation on several matrix ligands requires engagement of, and signalling via, a syndecan co-receptor. In this respect, syndecan-4 is the best-characterised family member, with its importance for migration *in vivo* being

Figure 1



Rab11 positive vesicles accumulate at the front of cells migrating in 3D. Ovarian carcinoma cells (A2780) transiently expressing Life-act GFP and mCherry-Rab11 were plated on cell-derived matrices for 6 hours in the presence of cyclic RGD<sub>N</sub> peptide and imaged using a spinning disk microscope. Maximum projections of z-stacks are displayed (scale bar = 10  $\mu\text{m}$ ). The 3D reconstruction was performed using the Imaris software. To allow the visualisation of the Rab11 positive vesicles, only the bottom half of the actin stack was displayed.

Figure 2



exemplified by the wound healing and angiogenesis defects observed in null mice [25]. In cells, cooperation between integrins and syndecan-4 has been demonstrated to regulate directional cell migration by dictating the spatiotemporal activation of the small GTPases Rac1, RhoA, RhoG and Arf6 [26–28,29\*,30\*\*].

Like integrins and growth factor receptors, syndecan function is regulated by endocytic trafficking [31\*,32]. Whilst it has been suggested that syndecans can be internalised by macropinocytosis [33\*], and that syndecan internalisation can be mediated via the binding of Rab5 to the syndecan-1 cytoplasmic domain [34], the mechanisms describing endocytosis of syndecans themselves are incompletely described. Syndecan recycling back to the plasma membrane has been shown to be dependent on a syndecan-syntenin-PIP2 association and the activity of the small GTPase Arf6 [32]. Interestingly, disruption of syndecan recycling by mutating the syntenin-PIP2 binding site triggers the accumulation of FGF receptor and  $\beta$ 1 integrin to syndecan-containing endosomes. These observations suggest that syndecans could participate in the recycling of adhesion and growth factor receptors, possibly by trapping receptors into a specific endosomal compartment through their glycosaminoglycan chains [32].

Recent studies have indicated that syndecans are more than just passive cargos trafficked to and from the plasma membrane. Indeed, the syndecan-syntenin interaction has been shown to promote the formation of exosomes by recruiting ALIX [35\*\*]. These data suggest that syndecans act as scaffolding platforms that recruit the machinery responsible for membrane budding and fission. Interestingly, exosome production was demonstrated to support tumour growth and metastasis, suggesting that syndecan functions could regulate these processes [36].

Syndecans have also been reported to regulate cell migration by controlling the internalisation and recycling of multiple receptors. Consistent with a regulatory role in receptor endocytosis, syndecans (in particular syndecan-4) have been shown to mediate the macropinocytosis of FGFR1 in response to FGF2 [33\*], the clathrin-dependent internalisation of Wnt-receptor in response to R-spondin in *Xenopus* [37] and caveolar endocytosis of  $\alpha$ 5 $\beta$ 1 integrin in response to H/0 (a soluble syndecan-4-binding fragment of FN) [29\*]. In this context, the syndecan-4-mediated endocytosis of  $\alpha$ 5 $\beta$ 1 has been shown to facilitate adhesion turnover and regulate directional cell

migration in 3D microenvironments and efficient wound healing *in vivo* [29\*].

In addition, syndecan-4 can dictate specificity of recycling between integrin heterodimers and therefore control the mode of cell migration [30\*\*]. Src-dependent phosphorylation of syndecan-4 was shown to suppress Arf6 activity and to promote the recycling of  $\alpha$ v $\beta$ 3 integrin to the cell surface, leading to adhesion stabilisation and rapid directional migration on 2D substrates [30\*\*]. Conversely, H/0-mediated stimulation of syndecan-4, or expression of a syndecan-4 construct that cannot be phosphorylated by Src, promoted Arf6 activation and the recycling of  $\alpha$ 5 $\beta$ 1 integrin, resulting in adhesion turn-over and random migration on 2D substrates [30\*\*]. Importantly, Src-mediated phosphorylation of syndecan-4 occurs in the conserved domain, present in all syndecan family members, and may represent a general mechanism whereby syndecans regulate receptor trafficking [30\*\*]. Interestingly, Src can be activated by various receptors including integrins and growth factor receptors [38–40], allowing potential feedback loops within the recycling pathway. Furthermore, this recent study supports previous observations relating to the heterodimer specific signalling and trafficking to promote cell migration in 2D and in 3D [18,19,20\*\*,41] (Figure 2).

Whether syndecans are involved in general receptor uptake or in the internalisation of specific receptors remains to be determined. As syndecans bind to an array of extracellular ligands, it will be important to assess whether specific syndecan ligands induce distinct internalisation pathways, or whether the internalisation route is dictated by the internalised receptor. A further priority will be to determine whether syndecans are internalised and trafficked together with, or separately from, these receptors.

From recent studies it is clear that syndecans, in particular syndecan-4, play a key role during cell migration by regulating the activation of various small GTPases and the trafficking of adhesion receptors. It remains to be elucidated whether these functions are independent or whether the syndecan-mediated temporal activation of small GTPases could be a consequence of the recycling pathway.

## Conclusion

Here, we have reviewed the recent advances that have altered perceptions of the role of adhesion receptor

**(Figure 2 Legend)** The mechanisms underlying the reciprocal nature of  $\alpha$ v $\beta$ 3 and  $\alpha$ 5 $\beta$ 1 recycling. In many cell types, including cancer cells and fibroblasts,  $\alpha$ v $\beta$ 3 recycling suppresses the recycling of  $\alpha$ 5 $\beta$ 1 to promote lamellipodial migration in 2D and invasion into ECM that lacks FN (**a**). Intervening in the recycling of  $\alpha$ v $\beta$ 3, by manipulating  $\alpha$ v $\beta$ 3 directly, expressing mutant p53 (in cancer cells), or by influencing syndecan phosphorylation/engagement promotes the recycling of  $\alpha$ 5 $\beta$ 1, and consequently a RhoA-ROCK dependent mode of random migration in 2D, and invasion into FN-rich ECM (**b**). The studies summarised above are persuasive of the notion that the signalling and trafficking events governed by adhesion receptors such as integrins and syndecans should be viewed as a network, rather than individual, isolated events. Red arrows delineate signalling events whilst black arrows indicate endocytic trafficking.



trafficking during cell movement. An emerging insight is the close connection between membrane dynamics and signalling, and we are beginning to clarify how these processes combine together to contribute to cell migration in a range of events *in vivo*. Whilst there is still much to be determined, some future perspectives can be discerned.

The approaches used to define the signalling events that are triggered by, and contribute to, receptor trafficking have in large part been defined by biochemical and immunocytochemical approaches. These techniques either lack precision or necessarily involve averaging of large cell populations. A priority for the future will therefore be improved precision, whether this involves localisation of signals to different membranes or pinpointing the sites at which vesicle budding and fusion occur.

A further priority will be to understand the variation in processes that underpin different modes of migration in different systems, and the mechanisms of switching that take place in relation to changes in cell phenotype and the mechanochemical environment of the cell. These studies will require a move to analysing ever more physiologically relevant, reconstituted 3D systems in which ECM composition, growth factors and mechanical properties have been reproduced, or the use of transparent organisms or intravital microscopy.

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## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Hynes RO: **Integrins: bidirectional, allosteric signaling machines.** *Cell* 2002, **125**:673-687.
  2. Wolfenson H, Lavelin I, Geiger B: **Dynamic regulation of the structure and functions of integrin adhesions.** *Dev Cell* 2013, **24**:447-458.
  3. Bridgewater RE, Norman JC, Caswell PT: **Integrin trafficking at a glance.** *J Cell Sci* 2012, **125**:3695-3701.
  4. Caswell PT, Norman JC: **Integrin trafficking and the control of cell migration.** *Traffic* 2006, **7**:14.
  5. Chao W-T, Kunz J: **Focal adhesion disassembly requires clathrin-dependent endocytosis of integrins.** *FEBS Lett* 2009, **583**:1337-1343.
  6. Ezratty EJ, Bertaux C, Marcantonio EE, Gundersen GG: **Clathrin mediates integrin endocytosis for focal adhesion disassembly in migrating cells.** *J Cell Biol* 2009, **187**:733-747.
  7. Nishimura T, Kaibuchi K: **Numb controls integrin endocytosis for directional cell migration with aPKC and PAR-3.** *Dev Cell* 2007, **13**:15-28.
  8. Ramsay AG, Keppler MD, Jazayeri M, Thomas GJ, Parsons M, Violette S, Weinreb P, Hart IR, Marshall JF: **HS1-associated protein X-1 regulates carcinoma cell migration and invasion via clathrin-mediated endocytosis of integrin alphavbeta6.** *Cancer Res* 2007, **67**:5275-5284.
  9. Steinberg F, Heesom KJ, Bass MD, Cullen PJ: **SNX17 protects integrins from degradation by sorting between lysosomal and recycling pathways.** *J Cell Biol* 2012, **197**:219-230.
  10. Böttcher RT, Stremmel C, Meves A, Meyer H, Widmaier M, Tseng H-Y, Fässler R: **Sorting nexin 17 prevents lysosomal degradation of  $\beta 1$  integrins by binding to the  $\beta 1$ -integrin tail.** *Nat Cell Biol* 2012, **14**:584-592.
  11. Krndjia D, Münzberg C, Maass U, Hafner M, Adler G, Kestler HA, Seufferlein T, Oswald F, Von Wichert G: **The phosphatase of regenerating liver 3 (PRL-3) promotes cell migration through Arf-activity-dependent stimulation of integrin  $\alpha 5$  recycling.** *J Cell Sci* 2012, **125**:3883-3892.
  12. Chen D-Y, Li M-Y, Wu S-Y, Lin Y-L, Tsai S-P, Lai P-L, Lin Y-T, Kuo J-C, Meng T-C, Chen G-C: **The Bro1-domain-containing protein Myopic/HDPTP coordinates with Rab4 to regulate cell adhesion and migration.** *J Cell Sci* 2012, **125**:4841-4852.
  13. Caswell P, Norman J: **Endocytic transport of integrins during cell migration and invasion.** *Trends Cell Biol* 2008, **18**:257-263.
  14. Caswell PT, Vadrevu S, Norman JC: **Integrins: masters and slaves of endocytic transport.** *Nat Rev Mol Cell Biol* 2009, **10**:843-853.
  15. Caswell PT, Spence HJ, Parsons M, White DP, Clark K, Cheng KW, Mills GB, Humphries MJ, Messent AJ, Anderson KI *et al.*: **Rab25 associates with  $\alpha 5 \beta 1$  integrin to promote invasive migration in 3D microenvironments.** *Dev Cell* 2007, **13**:496-510.
  16. Caswell PT, Chan M, Lindsay AJ, McCaffrey MW, Boettiger D, Norman JC: **Rab-coupling protein coordinates recycling of  $\alpha 5 \beta 1$  integrin and EGFR1 to promote cell migration in 3D microenvironments.** *J Cell Biol* 2008, **183**:143-155.
  17. Rainero E, Caswell PT, Muller PAJ, Grindlay J, McCaffrey MW, Zhang Q, Wakelam MJO, Voudsen KH, Graziani A, Norman JC: **Diacylglycerol kinase  $\alpha$  controls RCP-dependent integrin trafficking to promote invasive migration.** *J Cell Biol* 2012, **196**:277-295.
- Whilst several studies have shown that trafficking of integrins can be localised to the cell front, this paper demonstrates that phospholipid signalling, in particular production of phosphatidic acid by DGK $\alpha$ , is key to the spatial regulation of integrin recycling.
18. Danen EHJ, Van Rheeën J, Franken W, Huvenneers S, Sonneveld P, Jalink K, Sonnenberg A: **Integrins control motile strategy through a Rho-cofilin pathway.** *J Cell Biol* 2005, **169**:515-526.
  19. White DP, Caswell PT, Norman JC:  **$\alpha v \beta 3$  and  $\alpha 5 \beta 1$  integrin recycling pathways dictate downstream Rho kinase signaling to regulate persistent cell migration.** *J Cell Biol* 2007, **177**:515-525.
  20. Christoforides C, Rainero E, Brown KK, Norman JC, Tokar A: **PKD controls  $\alpha v \beta 3$  integrin recycling and tumor cell invasive migration through its substrate Rabaptin-5.** *Dev Cell* 2012, **23**:560-572.
- This paper shows that phosphorylation of Rabaptin-5 by PKD controls  $\alpha v \beta 3$  recycling, which in turn suppresses recycling of  $\alpha 5 \beta 1$ . Furthermore, the authors show that  $\alpha v \beta 3$  recycling promotes invasion into ECM that lacks fibronectin, but inhibits invasion into fibronectin-rich ECM, indicating that observations regarding heterodimer-specific control of migration in 2D are relevant in 3D.
21. Muller PAJ, Caswell PT, Doyle B, Iwanicki MP, Tan EH, Karim S, Lukashchuk N, Gillespie DA, Ludwig RL, Gosselin P *et al.*: **Mutant p53 drives invasion by promoting integrin recycling.** *Cell* 2009, **139**:1327-1341.
  22. Muller PAJ, Trinidad AG, Timpson P, Morton JP, Zanivan S, Van den Berghe PVE, Nixon C, Karim SA, Caswell PT, Noll JE *et al.*: **Mutant p53 enhances MET trafficking and signalling to drive cell scattering and invasion [Internet].** *Oncogene* 2012 <http://dx.doi.org/10.1038/onc.2012.148>.
  23. Cheng KW, Lahad JP, Kuo W-L, Lapuk A, Yamada K, Auersperg N, Liu J, Smith-McCune K, Lu KH, Fishman D *et al.*: **The RAB25 small**

**GTPase determines aggressiveness of ovarian and breast cancers.** *Nat Med* 2004, **10**:1251-1256.

24. Dozyńkiewicz MA, Jamieson NB, Macpherson I, Grindlay J, Van den Berghe PVE, Von Thun A, Morton JP, Gourley C, Timpson P, Nixon C *et al.*: **Rab25 and CLIC3 collaborate to promote integrin recycling from late endosomes/lysosomes and drive cancer progression.** *Dev Cell* 2012, **22**:131-145.

This paper demonstrates that, in cancer cells, endocytosed integrins are sorted for recycling to the front or to the rear of the cell dependent on their activation status. In this way, trafficking of unligated integrins is spatially restricted at the cell front, whilst active integrins are trafficked towards the rear and recycled in this subcellular region to coordinate protrusion at the front with retraction of the rear.

25. Morgan MR, Humphries MJ, Bass MD: **Synergistic control of cell adhesion by integrins and syndecans.** *Nat Rev Mol Cell Biol* 2007, **8**:957-969.
26. Dovas A, Yoneda A, Couchman JR: **PKC $\beta$ -dependent activation of RhoA by syndecan-4 during focal adhesion formation.** *J Cell Sci* 2006, **119**:2837-2846.
27. Bass MD, Roach KA, Morgan MR, Mostafavi-Pour Z, Schoen T, Muramatsu T, Mayer U, Ballestrem C, Spatz JP, Humphries MJ: **Syndecan-4-dependent Rac1 regulation determines directional migration in response to the extracellular matrix.** *J Cell Biol* 2007, **177**:527-538.
28. Bass MD, Morgan MR, Roach Ka, Settleman J, Goryachev AB, Humphries MJ: **p190RhoGAP is the convergence point of adhesion signals from  $\alpha 5 \beta 1$  integrin and syndecan-4.** *J Cell Biol* 2008, **181**:1013-1026.
29. Bass MD, Williamson RC, Nunan RD, Humphries JD, Byron A, Morgan MR, Martin P, Humphries MJ: **A syndecan-4 hair trigger initiates wound healing through caveolin- and RhoG-regulated integrin endocytosis.** *Dev Cell* 2011, **21**:681-693.

Here the authors describe that H<sub>2</sub>O-mediated stimulation of syndecan-4 activates the small GTPase RhoG resulting in rapid internalisation of  $\beta 1$  integrin. Subsequently, the authors demonstrate that this mechanism is required for directional cell migration and efficient wound healing *in vivo*.

30. Morgan MR, Hamidi H, Bass MD, Warwood S, Ballestrem C, Humphries MJ: **Syndecan-4 phosphorylation is a control point for integrin recycling.** *Dev Cell* 2013, **24**:472-485.

This work demonstrates that src-mediated phosphorylation of syndecan-4 determines the mode of cell migration in 2D by dictating specificity of recycling between the integrin heterodimers  $\alpha 5 \beta 1$  and  $\alpha v \beta 3$ . The authors also elucidate the molecular mechanism by showing that syndecan-4 phosphorylation differentially regulates Arf6 activity and syntenin binding.

31. Lambaerts K, Van Dyck S, Mortier E, Ivarsson Y, Degeest G, Luyten A, Vermeiren E, Peers B, David G, Zimmermann P: **Syntenin, a syndecan adaptor and an Arf6 phosphatidylinositol 4,5-bisphosphate effector, is essential for epiboly and gastrulation cell movements in zebrafish.** *J Cell Sci* 2012, **125**:1129-1140.

This work demonstrates that the syndecan-syntenin-Arf6 pathway is required for efficient cell migration *in vivo*.

32. Zimmermann P, Zhang Z, Degeest G, Mortier E, Leenaerts I, Coomans C, Schulz J, N'Kuli F, Courtoy PJ, David G: **Syndecan recycling is controlled by syntenin-PIP2 interaction and Arf6.** *Dev Cell* 2005, **9**:377-388.

33. Eifenbein A, Lanahan A, Zhou TX, Yamasaki A, Tkachenko E, Matsuda M, Simons M: **Syndecan 4 regulates FGFR1 signaling in endothelial cells by directing macropinocytosis.** *Sci Signal* 2012, **5**:ra36.

This work demonstrates that syndecan-4 regulates FGFR1 macropinocytosis in a RhoG and Rab5-dependent manner in endothelial cells. Furthermore, the authors show that syndecan-4-mediated internalisation of FGFR1 modulates the kinetics of MAPK activation upon FGF2 stimulation.

34. Hayashida K, Stahl PD, Park PW: **Syndecan-1 ectodomain shedding is regulated by the small GTPase Rab5.** *J Biol Chem* 2008, **283**:35435.

35. Baietti MF, Zhang Z, Mortier E, Melchior A, Degeest G, Geeraerts A, Ivarsson Y, Depoortere F, Coomans C, Vermeiren E *et al.*: **Syndecan-syntenin-ALIX regulates the biogenesis of exosomes.** *Nat Cell Biol* 2012, **14**:677-685.

Exosomes are important mediators of cell signalling and are also emerging as critical factors in progression and development of different disease states. This work demonstrates that syndecans regulate exosome formation and membrane budding via the recruitment of the syntenin-ALIX complex.

36. Peinado H, Alečković M, Lavotshkin S, Matei I, Costa-Silva B, Moreno-Bueno G, Hergueta-Redondo M, Williams C, Garcia-Santos G, Ghajar C *et al.*: **Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET.** *Nat Med* 2012, **18**:883-891.

37. Ohkawara B, Glinka A, Niehrs C: **Rspo3 binds Syndecan 4 and induces Wnt/PCP signaling via clathrin-mediated endocytosis to promote morphogenesis.** *Dev Cell* 2011, **20**:303-314.

38. Brunton VG, MacPherson IRJ, Frame MC: **Cell adhesion receptors, tyrosine kinases and actin modulators: a complex three-way circuitry.** *Biochim Biophys Acta* 2004, **1692**:121-144.

39. Playford MP, Schaller MD: **The interplay between Src and integrins in normal and tumor biology.** *Oncogene* 2004, **23**:7928-7946.

40. Streuli CH, Akhtar N: **Signal co-operation between integrins and other receptor systems.** *Biochem J* 2009, **418**:491-506.

41. Danen EHJ, Sonneveld P, Brakebusch C, Fassler R, Sonnenberg A: **The fibronectin-binding integrins  $\alpha 5 \beta 1$  and  $\alpha v \beta 3$  differentially modulate RhoA-GTP loading, organization of cell matrix adhesions, and fibronectin fibrillogenesis.** *J Cell Biol* 2002, **159**:1071-1086.